

An Analogue System Displaying All the Important Processes of the Catalytic Cycles Involving Monooxomolybdenum(VI) and Desoxomolybdenum(IV) Centers

Victor N. Nemykin,[†] Scott R. Davie,[†] Sujit Mondal,[†] Nick Rubie,[‡] Martin L. Kirk,^{*,‡} Arpad Somogyi,§ and Partha Basu^{*,†}

The Department of Chemistry and Biochemistry, Duquesne University, Pittsburgh, Pennsylvania 15282, and The Department of Chemistry, The University of Arizona, Tucson, Arizona 85721, and

The Department of Chemistry, The University of New Mexico, Albuquerque, New Mexico 87131-1096.

Received September 27, 2001

A majority of the pyranopterin-containing molybdoenzymes catalyze the formal transfer of an oxygen atom between substrate and solvent water,^{1,2} and these enzymes take part in the global nitrogen, sulfur, and arsenic cycles. A long-standing hypothesis states that the Mo active sites shuttle between dioxomolybdenum(VI) ([Mo^{VI}O₂]²⁺) and monooxomolybdenum(IV) ([Mo^{IV}O]²⁺) centers during the course of catalysis. Studies in several laboratories^{3,4,5} have demonstrated that a variety of discrete oxomolybdenum complexes can carry out this chemistry when reacted with oxygen atom abstractors such as tertiary phosphines. Although, tertiary phosphines are not physiological substrates, they are the most commonly used substrates in for studying oxygen atom transfer (OAT) reactions in model inorganic systems. Interestingly, using ¹⁸O-labeling it has been convincingly demonstrated that the DMSO reductase (DMSOR) from Rhodobacter sphaeroides can catalyze the transfer of an oxygen atom from DMSO to a water-soluble tertiary phosphine.⁶ The subsequent crystal structures,^{7,8} EXAFS⁹ and resonance Raman studies¹⁰ on the R. sphaeroides enzyme were interpreted in terms of monooxomolybdenum(VI) ([MoVIO]4+) and desoxomolybdenum(IV) ([Mo^{IV}]⁴⁺) centers, respectively. The oxidized [Mo^{VI}O]⁴⁺ center regains the catalytic competency by coupled electron proton-transfer processes. Similar structure-based reaction mechanisms have also been proposed for dissimilatory nitrate reductase11 and arsenite oxidase.12 Importantly, small molecule OAT reactivity in [MoVIO]4+ centers are less common,13,14 and CEPT reactions involving this center or [Mo^{IV}]4+ are unknown. Furthermore, in all previous cases of OAT reactivity involving discrete [MoVIO]4+ centers, the metal center is coordinated by sulfur donor ligands. This implies that Mo-S coordination may be essential for such reactivity. Here we report an example of OAT reactivity in a sixcoordinate monooxo Mo(VI) compound which does not possess sulfur donor ligands. In addition, we report the isolation of the intermediate in the OAT reaction and the regeneration of the oxo Mo(V) center using water as a source of the terminal oxygen. To our knowledge, this is the first demonstration of OAT and CEPT reactions involving discrete monooxo Mo(VI) and desoxo Mo(IV) centers.

A six-coordinate monooxo Mo(VI) compound was synthesized from the corresponding Mo(V) complex, LMo^VO(p-O-C₆H₄- $OC_2H_5_2$ (1), where L^- = hydrotris (3,5-dimethyl-1-pyrazolylborate). Compound 1 exhibits reversible one-electron oxidative and reductive couples.¹⁵ Bulk electrolysis confirmed the one-electron nature of the oxidative process and demonstrates that the cationic $[LMo^{VI}O(p-O-C_6H_4-OC_2H_5)_2]^+$ ([2]) species is stable enough to



Figure 1. Positive ESIMS spectra of [2]NO₃ in MeCN (A) before, (B) after addition of PPh₃, and (C) isolated [3]NO₃.

be isolated. This dark colored cationic species has been synthesized by chemical oxidation with (NH₄)₂Ce(NO₃)₆ and isolated as the nitrate salt [2]NO₃ in very good yields (74%). Solutions of the blue compound degrade to intractable products upon prolonged standing. Although, the instability of solutions of [2]NO₃ has frustrated attempts at obtaining single crystals, the complex has been thoroughly characterized by a variety of spectroscopic techniques. The ¹H NMR spectrum of [2]NO₃ is consistent with a six-coordinate species, and the IR spectrum exhibits a strong Mo=O stretch at 939 cm⁻¹. Acetonitrile solutions of the dark blue solid display strong absorptions at 850 and 575 nm assignable as phenolate to Mo charge-transfer transitions.

Positive electrospray ionization mass spectra (ESI-MS) of MeCN solutions of [2]NO₃ show the isotope pattern characteristic for the molecular ion around m/z 685 (C₃₁H₄₀BO₅N₆Mo) (Figure 1a), affirming the composition of the complex. The positive FAB mass spectrum of [2] shows a characteristic isotope pattern of the molecular ion peaks in the region m/z 685 and a fragmentation peak around m/z 546 associated with loss of a phenolic ligand. The deep blue color of 2 in acetonitrile is bleached by addition of tertiary phosphines such as PMe₃, PEt₃, PPh₃, and PEtPh₂, and the product of the PPh₃ reaction has been investigated by mass spectrometry. This product displays new peaks centered at m/z 669. Additionally, a peak for OPPh₃ is also present in the FAB ($[M + H]^+$), ESI ([M $(+ H]^+$ and EI (M^{•+}) mass spectra. The isotope distribution agrees well with that expected for the $[LMo^{IV}(p-O-C_6H_4-OC_2H_5)_2]^+$ ([3]) ion, consistent with the loss of an O atom from [2]. High-resolution

^{*} Author for correspondence. E-mail: basu@duq.edu.

Duquesne University.
 University of New Mexico.
 The University of Arizona.



accurate mass measurements have been used to confirm the elemental composition of both [3] ($C_{31}H_{40}BO_4N_6Mo$) and [2] ($C_{31}H_{40}BO_5N_6Mo$). It is important to note that the peaks associated with [3] (base peak m/z 669) appear in the positive ESI mass spectrum (Figure 1) in the absence of added acid, affirming the cationic nature of the new desoxo Mo species. A control experiment involving neutral 1 and PPh₃ did not show peaks around m/z 669. Also, the mass spectra of the reaction mixtures (with PMe₃, PEt₃, PPh₃, and PEtPh₂) exhibit peak clusters corresponding to the phosphine oxide adduct. The mass spectra suggest that the OAT reaction from monooxo Mo(VI) centers proceeds through phosphineoxide-bound intermediate similar to the intermediate described for dioxo Mo(VI) centers.¹⁶ For reaction with PPh₃ this adduct, [LMo(OPPh₃)(p-O-C₆H₄ $-OC_2H_5$)₂]⁺NO₃⁻, has been isolated and spectroscopically characterized.

The brown residue obtained from reacting $[2]NO_3$ with PPh₃ has been purified by column chromatography on silica gel using acetonitrile as an eluent. Evaporation of the first brown band yielded a brown compound whose positive ion ESI mass spectrum exhibits a strong peak centered at 669 and the isotope distribution pattern agrees well with the solution study undertaken for $[3]NO_3$ (Figure 1C).¹⁷ The isolated product displays no characteristic Mo=O stretch, confirming the desoxo nature of $[3]NO_3$. The putative intermediate, $[LMo(OPPh_3)(p-O-C_6H_4OC_2H_5)_2]^+NO_3^-$, elutes as the second brown band from the same column. This compound exhibits characteristic mass spectrum, and no Mo=O vibration. The complex is further characterized by ¹H and ³¹P NMR spectroscopies. The OAT reaction was further probed by resonance Raman (rR) spectroscopy.¹⁸ The rR spectra of [2]NO₃ and the brown residue obtained by reacting [2]NO3 with PPh3 exhibit no Mo=O vibration (Supporting Information) indicating the Mo=O stretch observed for [2]NO₃ is lost upon addition of PPh₃.

The Mo=O vibration in the isotopically labeled $LMo^{V18}O(p-O C_6H_4$ -OC₂ H_5)₂ shifts 40 cm⁻¹ to lower frequency; a similar shift is also observed for Mo(VI) complex. When [LMo^{VI 18}O(p-O- $C_6H_4-OC_2H_5)_2]^+NO_3^-$ is reacted with PPh₃, peaks due to ¹⁸OPPh₃ and [LMo(¹⁸OPPh₃)(p-O-C₆H₄-OC₂H₅)₂]⁺ could be detected by ESI-MS.¹⁹ These findings clearly demonstrates that the terminal oxo group in [2]NO₃ has been transferred to PPh₃. When [3]NO₃ was reacted with H₂[¹⁸O] in the presence of 2,3-dicyano-5,6dichloro-1,4-benzoquinone (DDQ), the resulting solution exhibit a peak cluster due to LMo^{V 18}O(p-O-C₆H₄-OC₂H₅)₂. Evaporation of the solution resulted a brown mass, whose IR spectrum is almost identical to that of authentic LMo^V ¹⁸O(p-O-C₆H₄-OC₂H₅)₂.²⁰ These experiments demonstrate that not only has the terminal oxo group of $[2]NO_3$ been transferred to PPh₃ but it can be regenerated from water. Thus, the analogue system displays all of the fundamental aspects of the catalytic cycle represented in Scheme 1.

In summary, we have demonstrated that a six-coordinate monooxo Mo(VI) center can carry out OAT chemistry *in the absence of thiolate (or ene-dithiolate) donor ligands and the monooxo Mo(VI) center can be regenerated from a physiologically relevant water molecule.* In addition, we have isolated a putative OAT intermediate *from a monooxo Mo(VI) center*. Thus, the chemistry described here parallels the previously proposed enzymatic reactions. These are important findings with respect to fully understanding the primary role of the pyranopterin ene-1,2-dithiolate in the oxidative and reductive half reactions of enzymes that cycle between monooxo Mo(VI) and desoxo Mo(IV) centers.

Acknowledgment. We are grateful to Professor John Enemark for numerous stimulating discussions. Financial support from the National Institute of Health (GM 615501 to P.B., and GM 057378 to M.L.K.), and Winters Foundation (to P.B.) is gratefully acknowledged.

Supporting Information Available: All synthetic procedures and characterization data for complexes, a table containing the results of accurate mass measurements for [2], [3], and OPPh₃, and Figures of rR spectra and IR spectra of isotopically labeled compounds (PDF). This material is available free of charge via the Internet at http:// pubs.acs.org.

References

- (a) Hille, R. Chem. Rev. **1996**, 96, 2757. (b) Pilato, R. S.; Stiefel, E. I. In Bioinorganic Catalysis; Reedjik, J.; Ed.; Marcel Dekker, Inc.: New York 1993; p 131. (c) Enemark, J. H.; Young, C. G. Adv. Inorg. Chem. **1993**, 32, 1.
- (2) Fischer, B.; Enemark, J. H.; Basu, P. J. Inorg. Biochem. 1998, 72, 13.
- (3) (a) Holm, R. H. Coord. Chem. Rev., 1990, 100, 183. (b) Schultz, B. E.; Gheller, S. F.; Muetterties, M. C.; Scott, M. J.; Holm, R. H. J. Am. Chem. Soc., 1993, 115, 2714. (c) Berg, J. M.; Holm, R. H. J. Am. Chem. Soc. 1985, 107, 925.
- (4) (a) Das, S. K.; Chaudhury, P. K.; Biswas, D.; Sarkar, S. J. Am. Chem. Soc. 1994, 116, 9061. (b) Dutta, S. K.; McConville, D. B.; Youngs, W. J.; Choudhury, M. Inorg. Chem. 1997, 36, 2517-2522. (c) Bhatterjee, S.; Bhattaryya, R. J. Chem. Soc., Dalton Trans. 1993, 1151. (d) Oku, H.; Ueyama, N.; Kondo, M.; Nakamura, A. Inorg. Chem. 1994, 33, 209.
- (5) (a) Laughlin, L. J.; Young, C. J. *Inorg. Chem.* **1996**, *35*, 1050. (b) Xiao, Z.; Bruck, M. A.; Enemark, J. H.; Young, C. Y.; Wedd, A. G. *Inorg. Chem.* **1996**, *35*, 7508. (c) Xiao, Z.; Young, C. G.; Enemark, J. H.; Wedd, A. G. J. Am. Chem. Soc. **1992**, *114*, 9194.
- (6) Schultz, B. E.; Hille, R.; Holm, R. H. J. Am. Chem. Soc. 1995, 117, 827.
 (7) (a) Schindelin, H.; Kiser, C.; Hilton, J.; Rajagopalan, K. V.; Rees, D. C. Science 1996, 272, 1615. (b) Li, H. K.; Temple, C.; Rajagopalan, K. V.;
- Schindelin, H. J. Am. Chem. Soc. 2000, 122, 7673.
 (8) The crystal structures of *R. capsulatus* protein exhibit a different coordination mode. Schneider, F.; Löwe, J.; Huber, R.; Schindelin, H.; Kiser, C.; Knäblein, J. J. Mol. Biol. 1996, 263, 53. McAlpine, A. S.; McEwan, A. G.; Bailey, S. J. Mol. Biol. 1998, 275, 613.
 (9) Correct C. N.; Witter T. T., C. D. D. C. D. S. C. S. S. McAlpine, M. S. S. McAlpine, A. S.; McEwan, S. S. Mol. Science, C. N.; Science, S. S. Mol. Biol. 1998, 275, 613.
- (9) George, G. N.; Hilton, J.; Temple,C.; Prince, R. C.; Rajagopalan, K. V. J. Am. Chem. Soc. 1999, 121, 1256.
- (10) Garton, S. D.; Hilton, J.; Oku, H.; Crousse, B. R.; Rajagopalan, K. V.; Johnson, M. K. J. Am. Chem. Soc. 1997, 119, 12906.
- (11) Dias, J. M.; Than, M. E.; Humm, A.; Huber, R.; Bourenkov, G. P.; Bartunik, H. D.; Bursakov, S.; Calvete, J.; Caldeira, J.; Carneiro, C.; Moura, J. J. G.; Moura, I.; Romão, M. J. *Structure* **1999**, *7*, 65.
- (12) Ellis, P. J.; Conrads, T.; Hille, R.; Kuhn, P. Structure 2001, 9, 125.
- (13) Arzoumanian, H.; Corao, C.; Krentzien, H.; Lopez, R.; Teruel, H. J. Chem. Soc., Chem Commun. 1992, 856
- (14) Donahue, J. P.; Lorber, C.; Nordlander, E.; Holm, R. H. J. Am. Chem. Soc. 1998, 120, 3259–3260. Lim, B. S.; Holm, R. H. J. Am. Chem. Soc. 2001, 123, 1920. Sung, K.-M.; Holm, R. H. J. Am. Chem. Soc. 2001, 123, 1931.
- (15) Chang, C-S. J.; Collision, D.; Mabbs, F. E.; Enemark, J. H. *Inorg. Chem.* **1990**, 29, 2261. McDonagh, A. M.; Ward, M. D.; McCleverty, J. A. *New J. Chem.* **2001**, 25, 1236.
- (16) Smith, P. D.; Millar, A. J.; Young, C. G.; Ghosh, A.; Basu, P. J. Am. Chem. Soc. 2000, 122, 9298.
- (17) The mass spectrum of the isolated desoxo compounds also exhibit a peak cluster ~m/z 710 due to the formation of a six-coordinate MeCN adduct. However, no characteristic stretch for nitrile could be identified in solidstate IR.
- (18) Solid-state rR spectra of [2]NO₃ and the brown residue obtained by reacting [2]NO₃ with PPh₃ were collected as a solids dispersed in an NaCl/NaNO³⁻ matrix. No Mo=O stretch was observed for the brown residue using 647.1, 514.5, 488.0, 457.9, or 413.1 nm excitation. For [2], excitation at 676 and 647 nm exhibit vibrations at 1150, 1247, 1493, and 1586 cm⁻¹, indicating coordinated phenol.
- (19) An OAT reaction has also been observed from [LMo^{VI}O(OMe)₂]⁺ Nemykin, V. and Basu, P. Unpublished results.
- (20) The estimated ¹⁸O-enrichment of [2]NO₃ (from direct synthesis) based on ESI MS is 38%, while it is 54% when prepared from [3]NO₃ with H₂¹⁸O/DDQ.

JA017178L